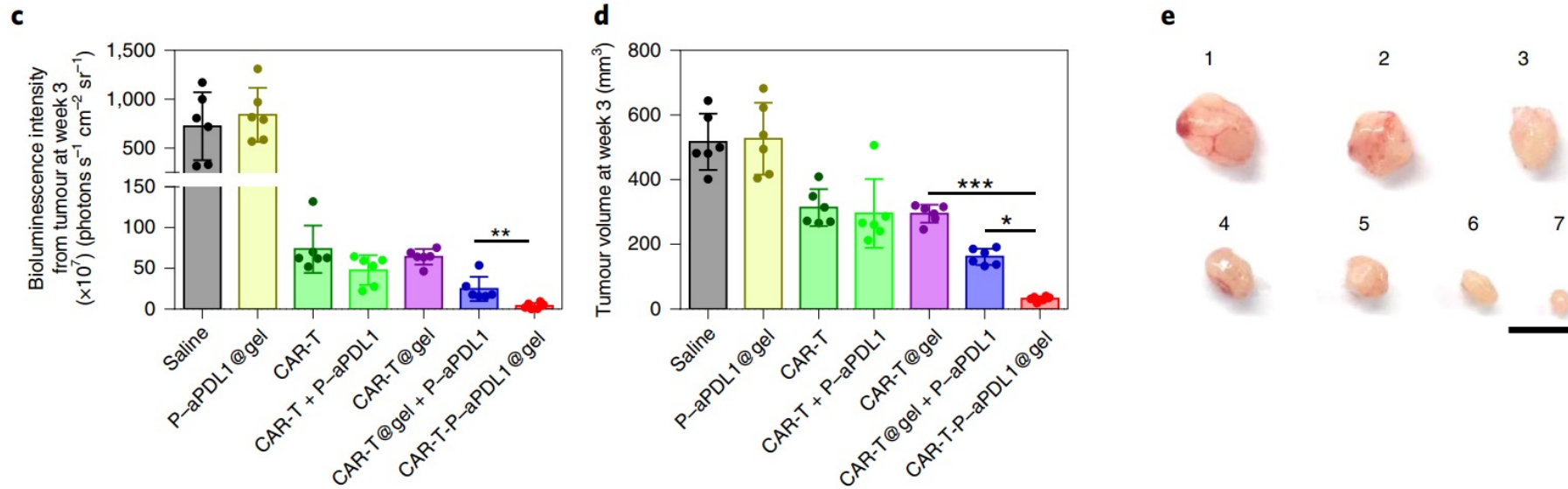
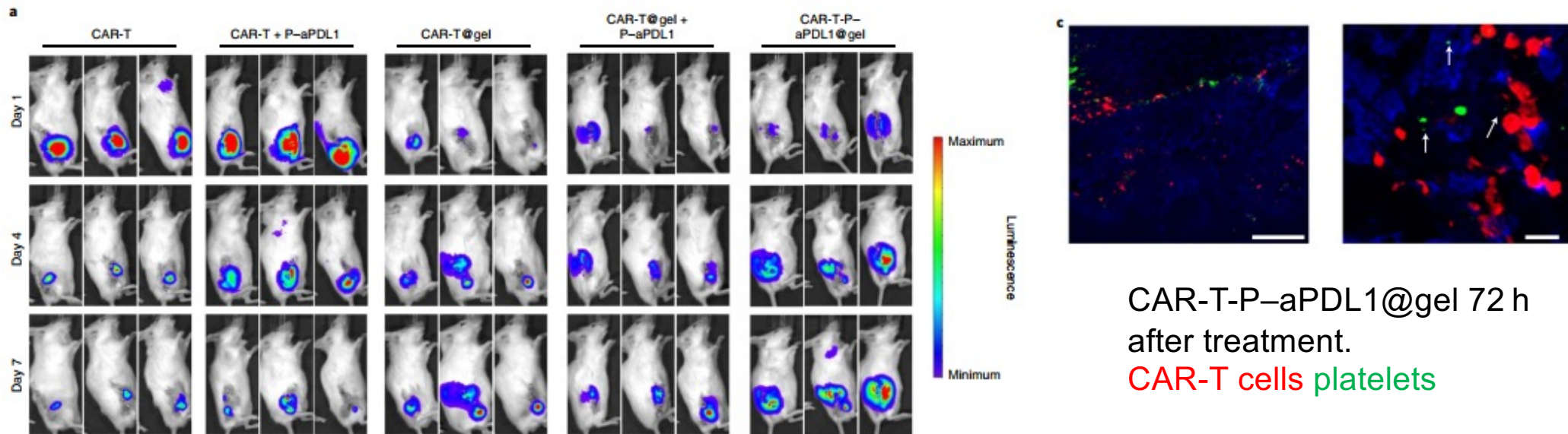


Inhibition of post-surgery tumour recurrence via a hydrogel releasing CAR-T cells and anti-PDL1-conjugated platelets

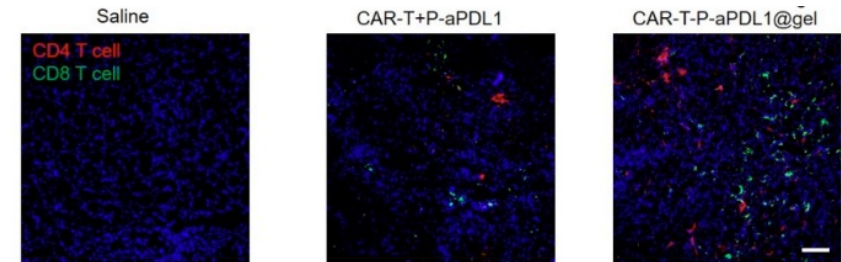
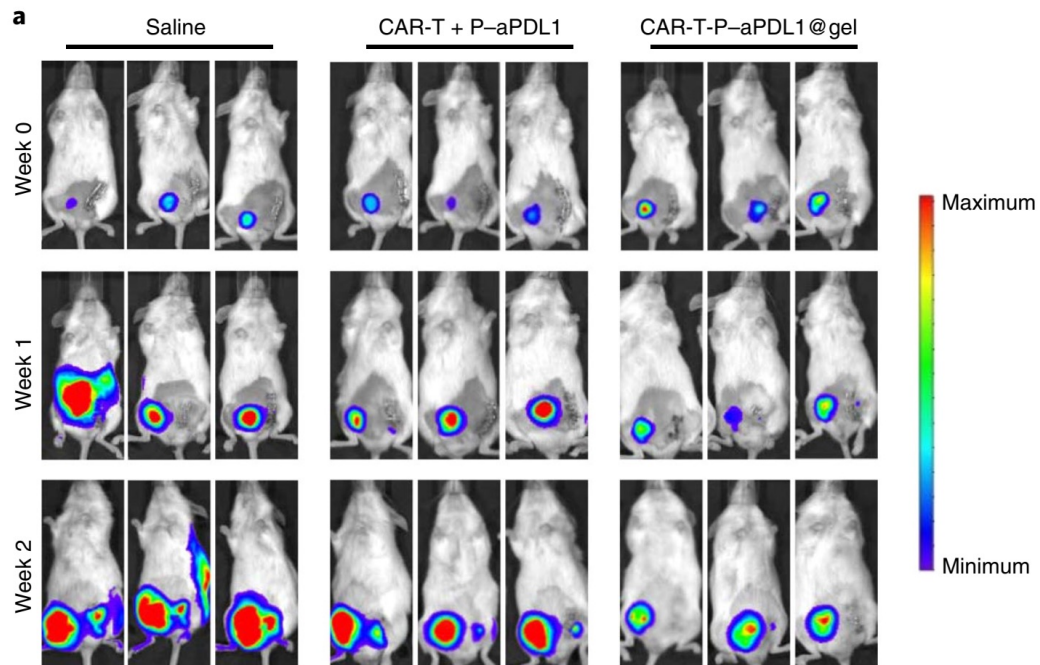


Inhibition of post-surgery tumour recurrence via a hydrogel releasing CAR-T cells and anti-PDL1-conjugated platelets



CAR-T cells encapsulated in the hydrogel persist and expand in vivo.

Inhibition of post-surgery tumour recurrence via a hydrogel releasing CAR-T cells and anti-PDL1-conjugated platelets



Representative confocal images of CD4 and CD8 T cells detected in the distant tumor.

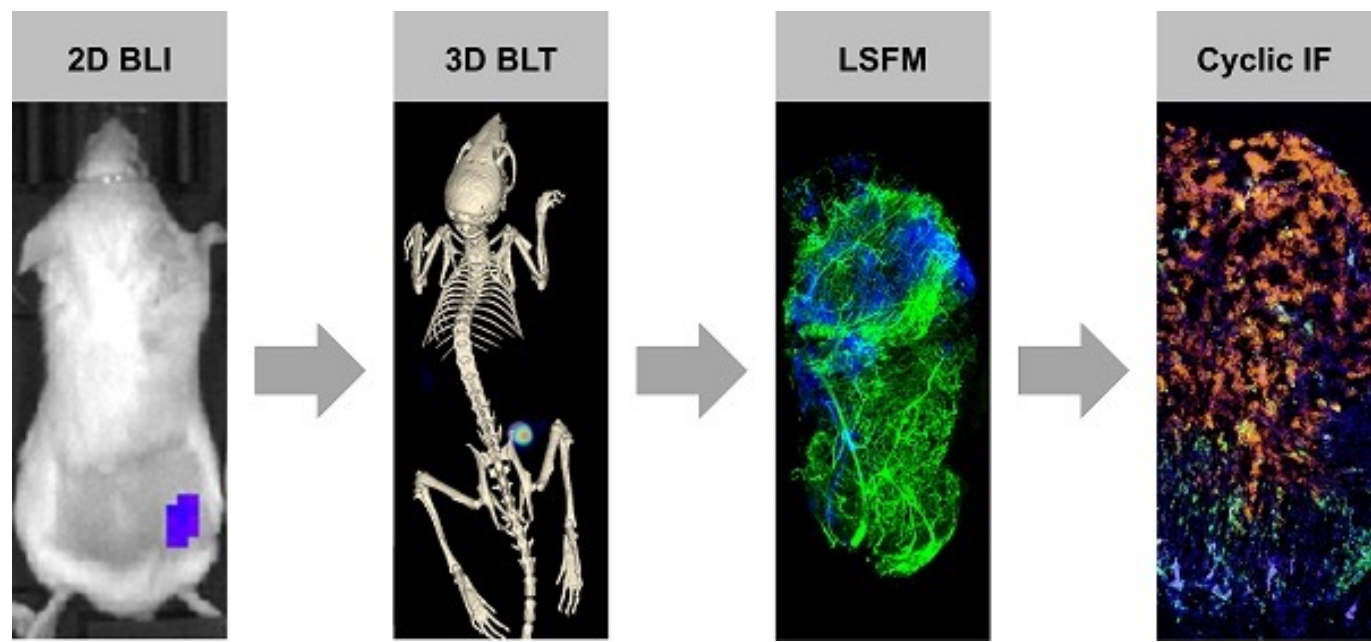
Engineered hydrogel cell delivery promotes abscopal antitumor effects

Multi-modal animal model

Research Paper

A multimodal imaging workflow for monitoring CAR T cell therapy against solid tumor from whole-body to single-cell level

Rita Pfeifer^{1*}, Janina Henze^{1,2*}, Katharina Wittich¹, Andre Gosselink^{1,3}, Ali Kinkhabwala¹, Felix Gremse⁴, Cathrin Bleilevens¹, Kevin Bigott¹, Melanie Jungblut¹, Olaf Hardt¹, Frauke Alves^{2,5}, Wa'el Al Rawashdeh^{1,6}✉



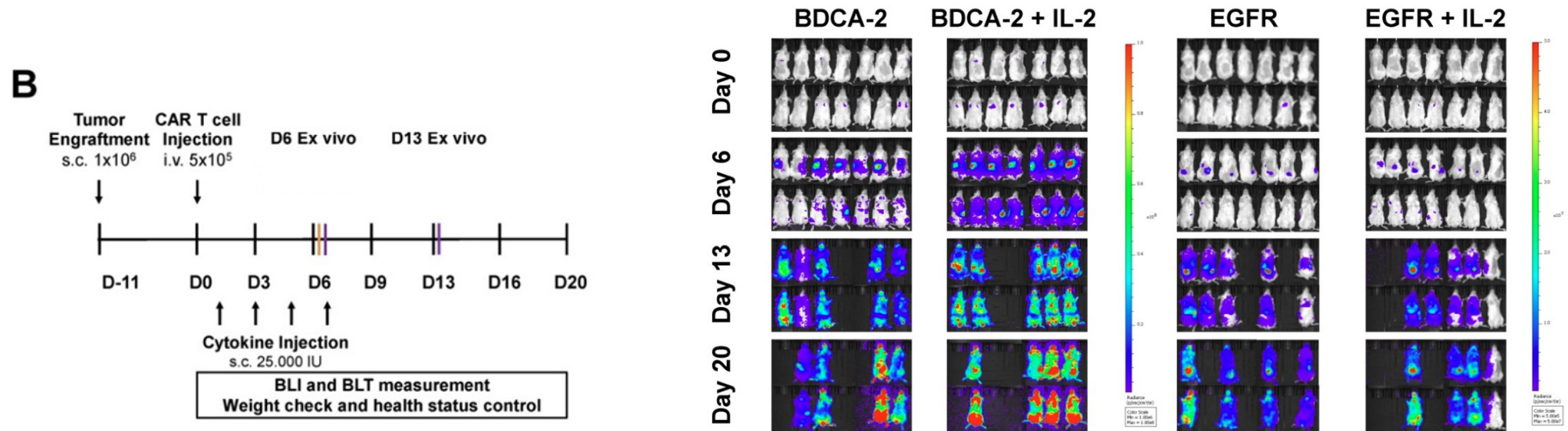
Theranostics

2022; 12(11): 4834-4850. doi: 10.7150/thno.68966

Research Paper

A multimodal imaging workflow for monitoring CAR T cell therapy against solid tumor from whole-body to single-cell level

Rita Pfeifer^{1*}, Janina Henze^{1,2*}, Katharina Wittich¹, Andre Gosselink^{1,3}, Ali Kinkhabwala¹, Felix Gremse⁴, Cathrin Bleilevens¹, Kevin Bigott¹, Melanie Jungblut¹, Olaf Hardt¹, Frauke Alves^{2,5}, Wa'el Al Rawashdeh^{1,6}

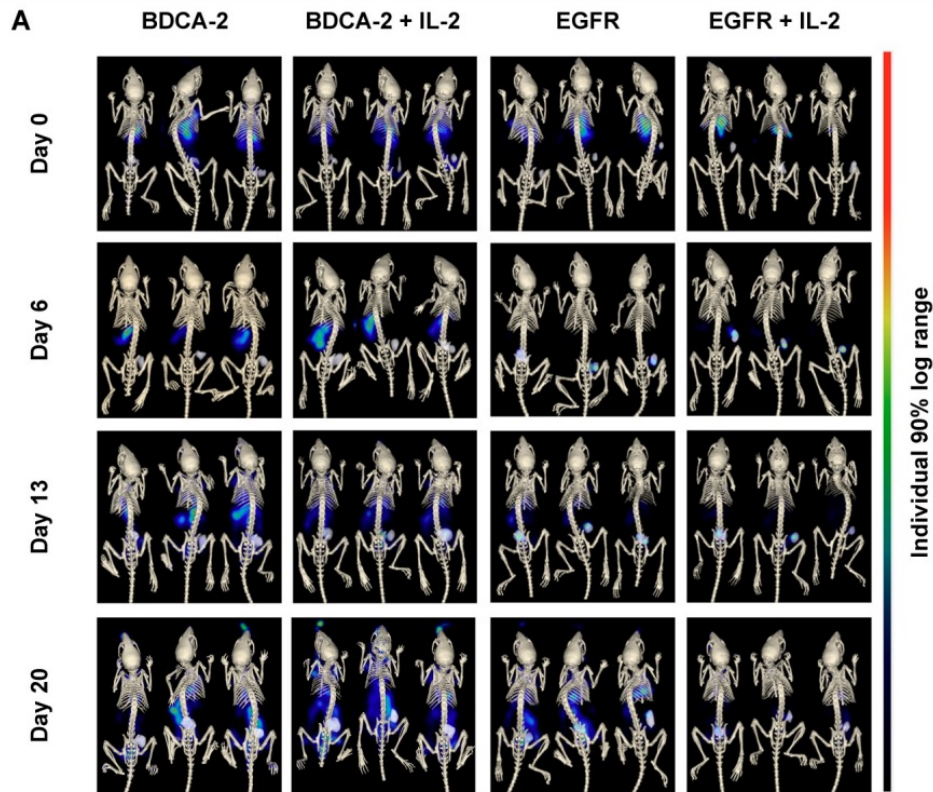


Therapeutic EGFR CAR T cells in comparison to control BDCA-2 CAR T cells in a subcutaneous pancreatic cancer xenograft mouse model

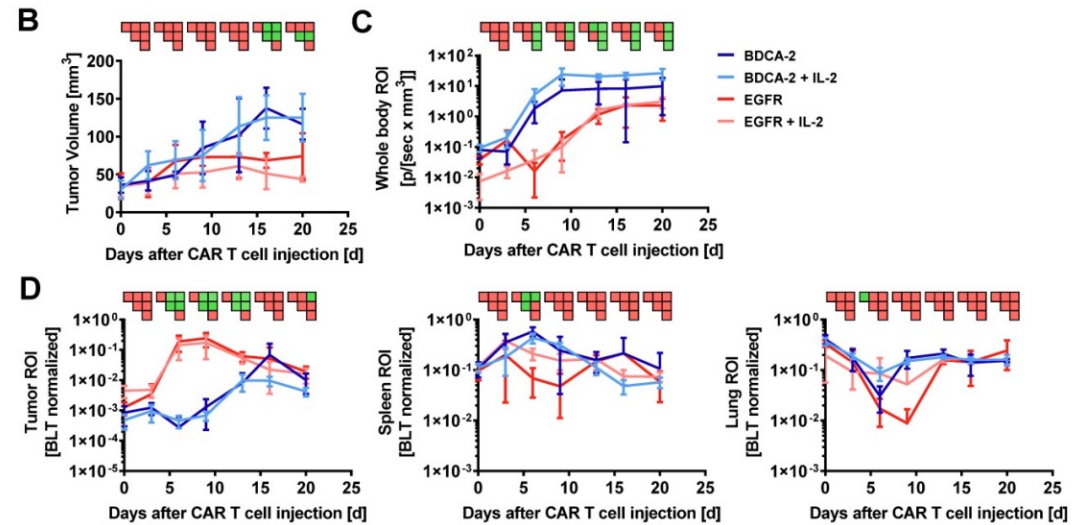
Research Paper

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Rita Pfeifer^{1*}, Janina Henze^{1,2*}, Katharina Wittich¹, Andre Gosselink^{1,3}, Ali Kinkhabwala¹, Felix Gremse⁴, Cathrin Bleilevens¹, Kevin Bigott¹, Melanie Jungblut¹, Olaf Hardt¹, Frauke Alves^{2,5}, Wa'el Al Rawashdeh^{1,6}



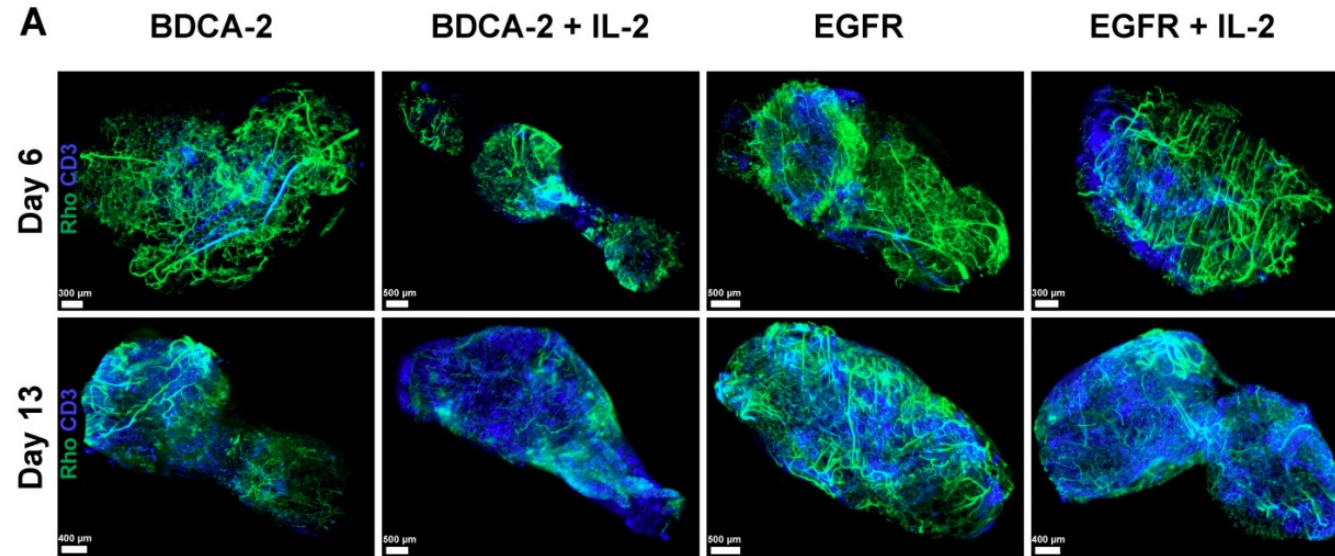
3D hybrid μ CT/BLT precisely detects antigen recognition-dependent intratumoral CAR T cell trafficking and expansion



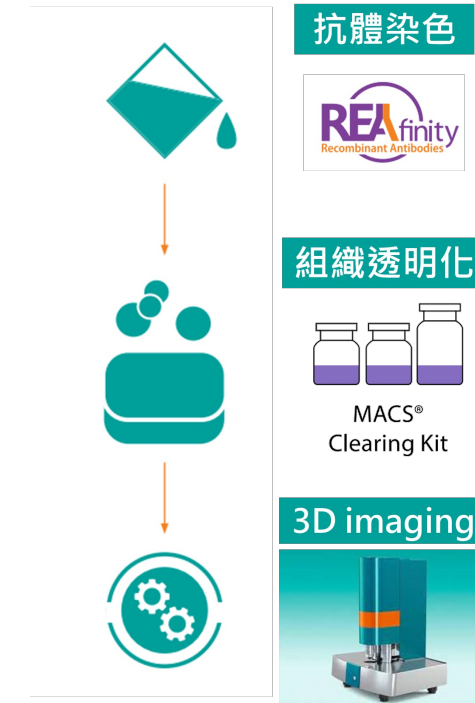
Research Paper

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LSFM Light sheet microscope



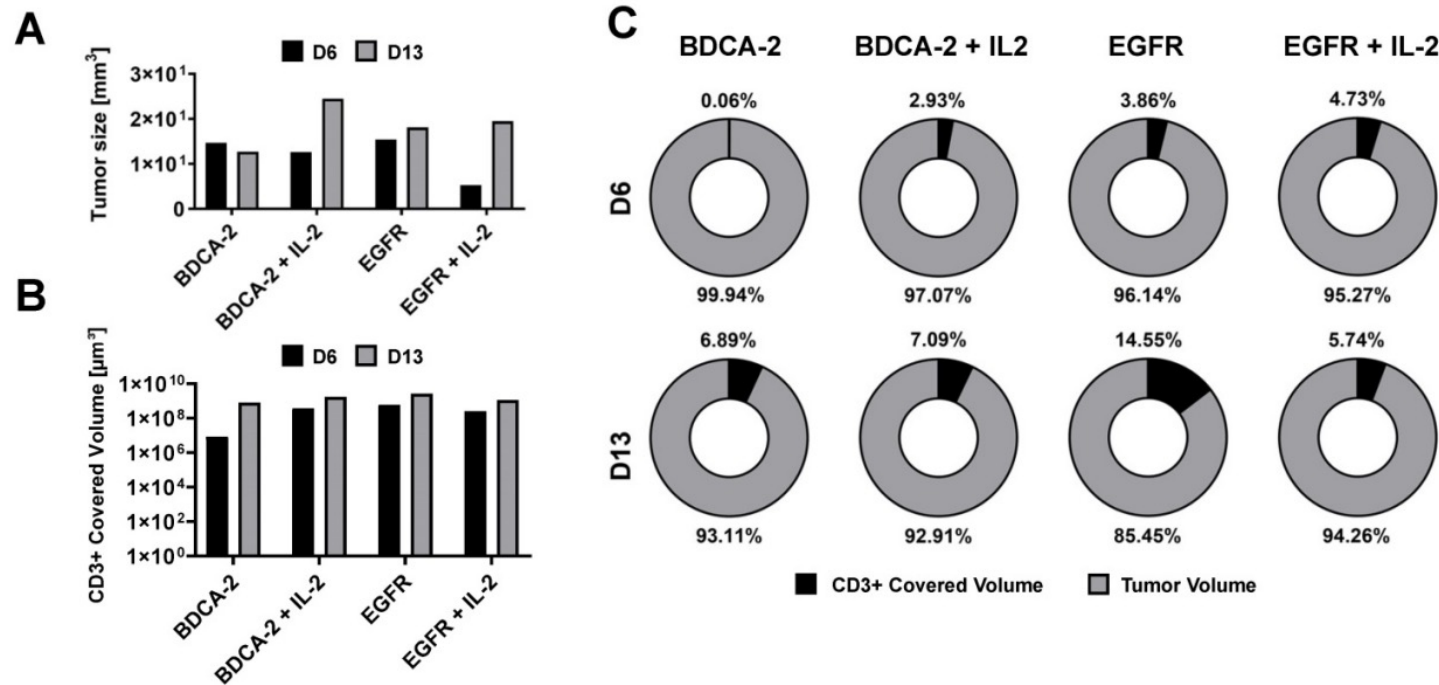
Theranostics

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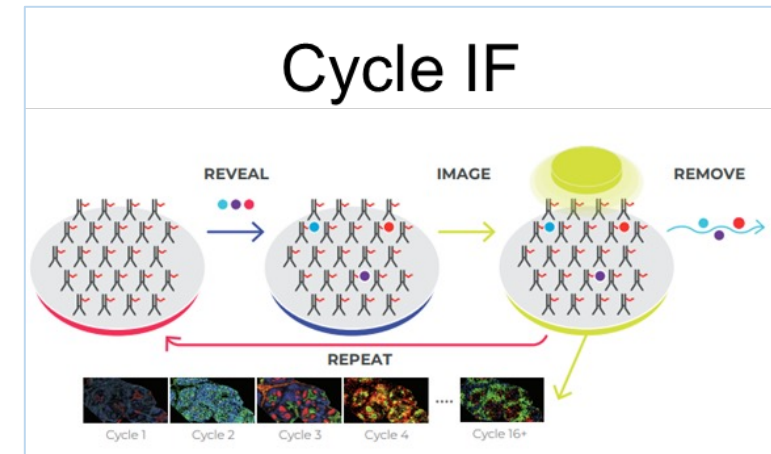
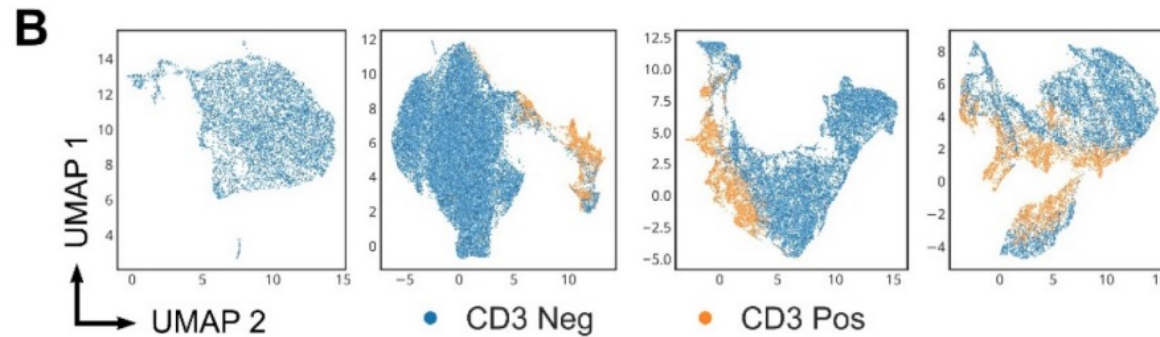
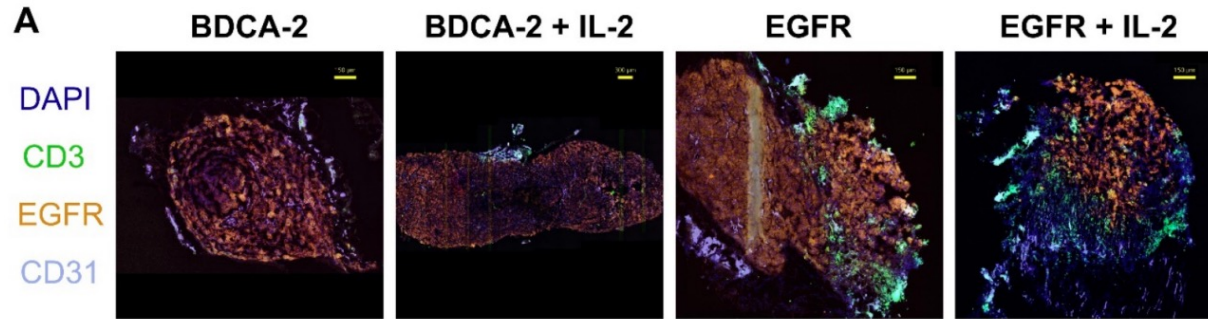


Spot-reconstruction analysis in LSM reveals effects of IL-2 on CAR T cells intratumoral distribution

Research Paper

A multimodal imaging workflow for monitoring CAR T cell therapy against solid tumor from whole-body to single-cell level


Rita Pfeifer^{1*}, Janina Henze^{1,2*}, Katharina Wittich¹, Andre Gosselink^{1,3}, Ali Kinkhabwala¹, Felix Gremse⁴, Cathrin Bleilevens¹, Kevin Bigott¹, Melanie Jungblut¹, Olaf Hardt¹, Frauke Alves^{2,5}, Wa'el Al Rawashdeh^{1,6}✉



High-content cyclic IF imaging reveals phenotypic changes in tumor and T cells following CAR T cell treatment

JnH total solution for Animal model

Theranostics 2022, Vol. 12, Issue 11 4834

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INTERNATIONAL PUBLISHER

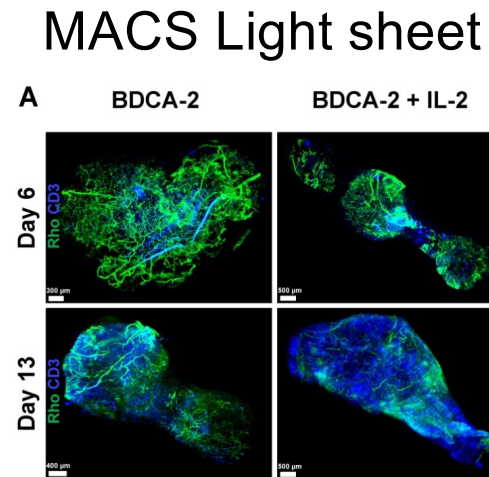
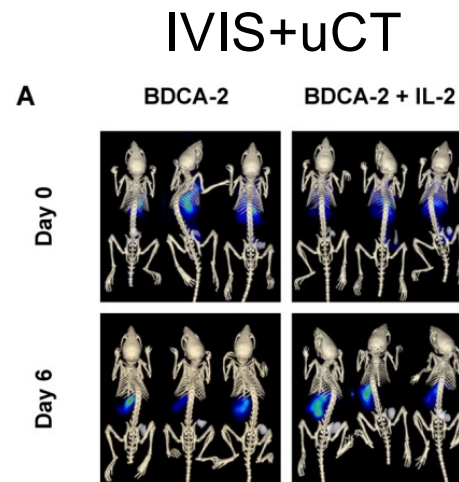
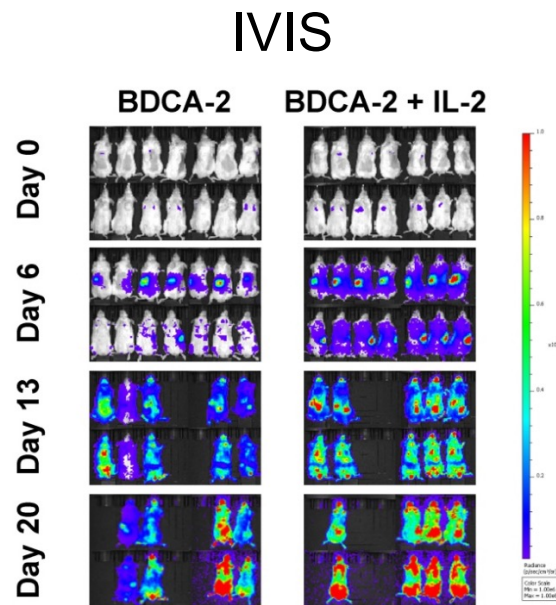
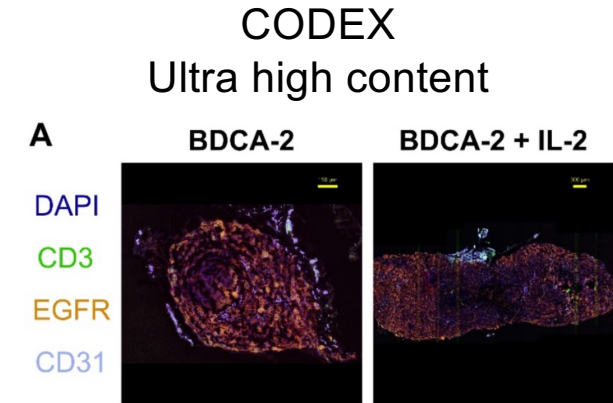
Theranostics

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Research Paper

A multimodal imaging workflow for monitoring CAR T cell therapy against solid tumor from whole-body to single-cell level

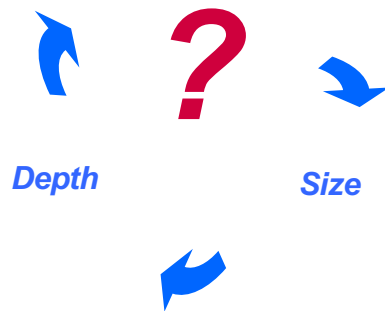
Rita Pfeifer^{1*}, Janina Henze^{1,2*}, Katharina Wittich¹, Andre Gosselink^{1,3}, Ali Kinkhabwala¹, Felix Gremse⁴, Cathrin Bleilevens¹, Kevin Bigott¹, Melanie Jungblut¹, Olaf Hardt¹, Frauke Alves^{2,5}, Wa'el Al Rawashdeh^{1,6}



Why 3D Optical Tomography? 2D VS 3D imaging



Concentration



Depth

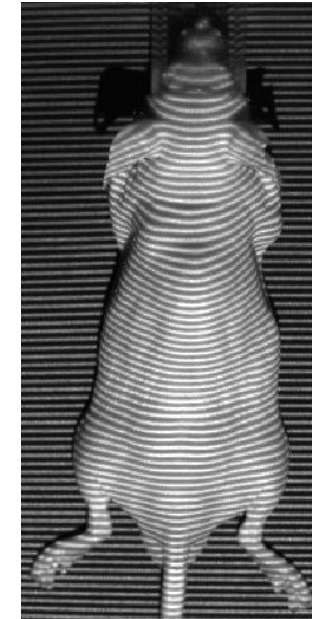
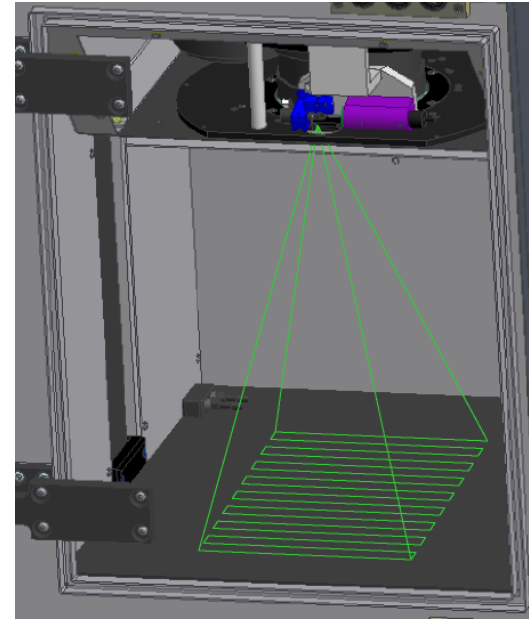
Size

- 2D成像獲得信號到達體表的相對強度
- 是否需要比較不同深度信號的強度？
3D成像能比較**不同深度**的信號強弱
- 是否要對信號進行定位和絕對定量？
3D成像能夠**定位**，還原信號的體積訊息，並且**絕對定量**

3D Imaging from a Single View is a Two-Step Process

- **Bioluminescence (Diffuse Light Imaging Tomography™):**

- Step 1: **Surface Topography**
- Step 2: Obtain images using **multiple emission filters** for **Tomographic** mapping of source location



3D Imaging from a Single View is a Two-Step Process

(12) **United States Patent**
Rice et al.

(10) **Patent No.:** US 8,044,996 B2
(45) **Date of Patent:** Oct. 25, 2011

(54) **SURFACE CONSTRUCTION USING
COMBINED PHOTOGRAPHIC AND
STRUCTURED LIGHT INFORMATION**

4,773,097 A	9/1988	Suzaki et al.
5,202,091 A	4/1993	Lisenbee
5,205,291 A	4/1993	Potter
5,242,441 A	9/1993	Avitall
6,216,200 A	6/2004	Mitchell et al.

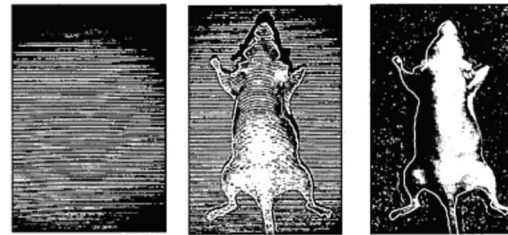
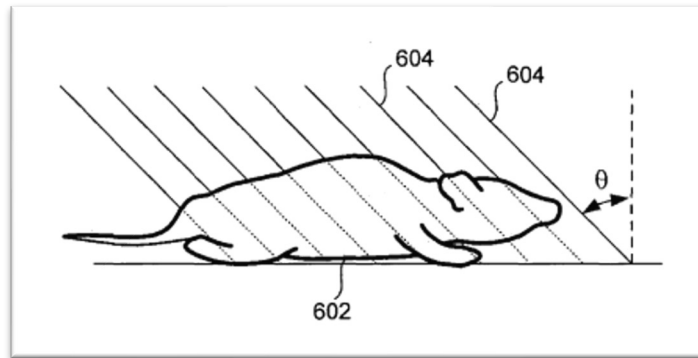


FIG. 8B FIG. 8C FIG. 8D

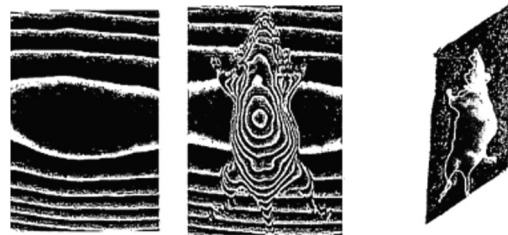
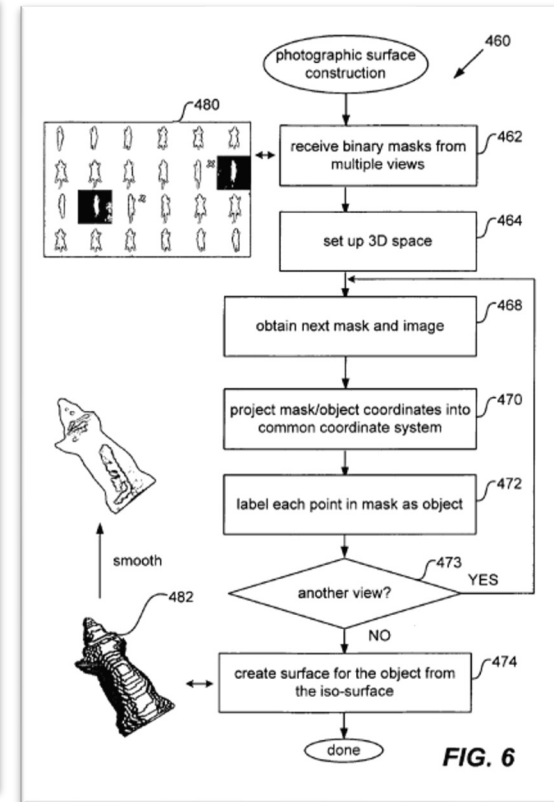


FIG. 8E FIG. 8F FIG. 8G



DLIT 3D Reconstruction

Analyze Properties Results

Tissue Properties: Mouse Tissue

Source Spectrum: Firefly

Plot: Tissue Properties

Luminescent Calibration :
Result not found

DLIT 3D Reconstruction

Analyze Properties Results

Sequence: EL20120517115255_SEQ
Tissue: Mouse Tissue Source: Firefly

Select Filters:

Filter	Threshold %
<input checked="" type="checkbox"/> 560	0.5
<input checked="" type="checkbox"/> 580	0.5
<input checked="" type="checkbox"/> 600	0.5
<input checked="" type="checkbox"/> 620	0.5
<input checked="" type="checkbox"/> 640	0.5

• Select wavelengths

NNLS Start

- Select tissue properties
- Select source spectrum

Data Adjustment

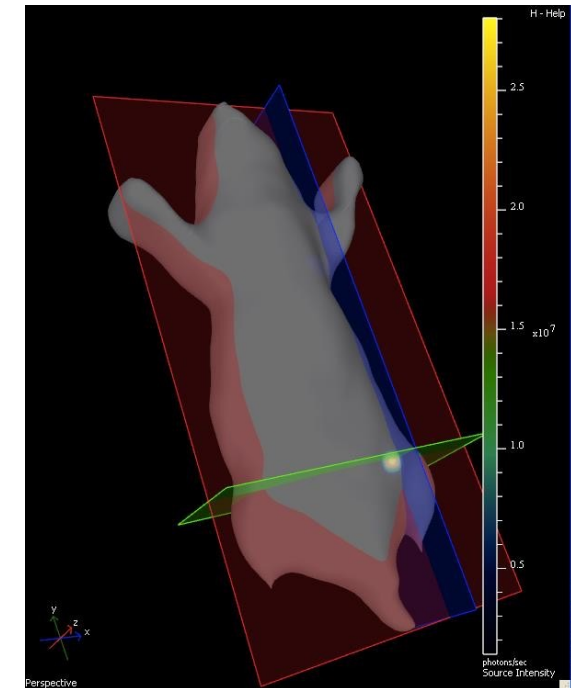
Threshold Tools
Adjustment: 0.5%
Min counts: 61

Region Selection Tools
Draw Erase
Painting size: Medium Segment: Red Opacity: [Slider]
Reset

Cancel Ok

- Threshold your data

3D reconstruction

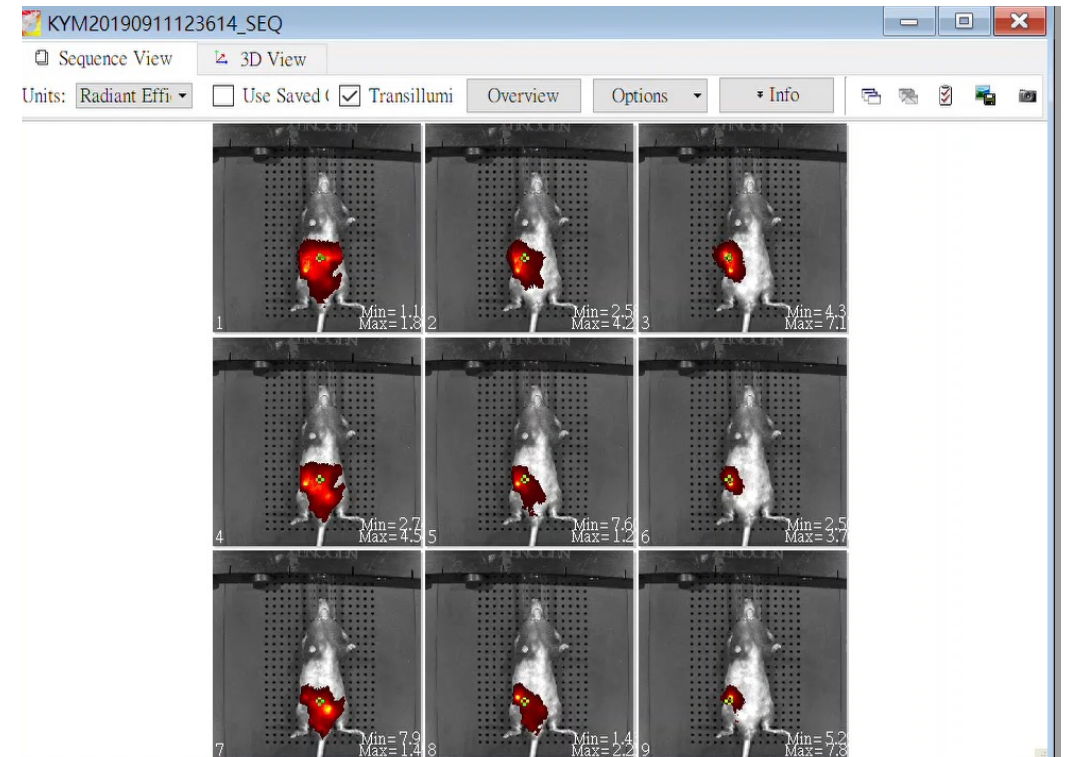
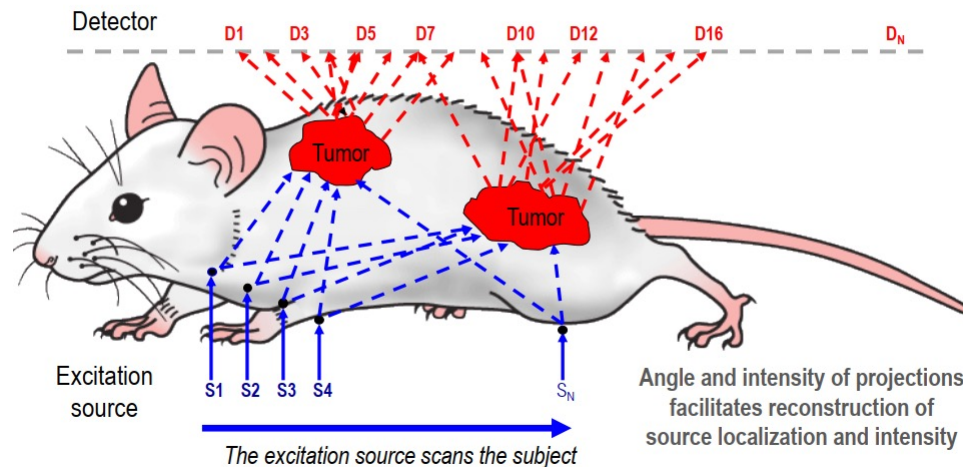


3D Imaging from a Single View is a Two-Step Process

● Fluorescence (Fluorescence Imaging Tomography):

- Step 1: Surface Topography
- Step 2: Obtain images using **multiple transillumination points**

The CCD collects multiple projections using designated emission filter



(12) **United States Patent**
Rice et al.

(10) **Patent No.:** US 7,599,731 B2
(45) **Date of Patent:** Oct. 6, 2009

(54) **FLUORESCENT LIGHT TOMOGRAPHY**

(75) **Inventors:** **Bradley W. Rice**, Danville, CA (US);
Chaincy Kuo, Oakland, CA (US);
Daniel G. Stearns, Mountain View, CA (US);
Heng Xu, Alameda, CA (US)

(73) **Assignee:** **Xenogen Corporation**, Alameda, CA (US)

4,773,097 A	9/1988	Suzaki et al.
5,202,091 A	4/1993	Lisenbee
5,205,291 A *	4/1993	Potter 600/431
5,242,441 A	9/1993	Avitall
5,319,209 A	6/1994	Miyakawa et al.
5,334,193 A	8/1994	Nardella

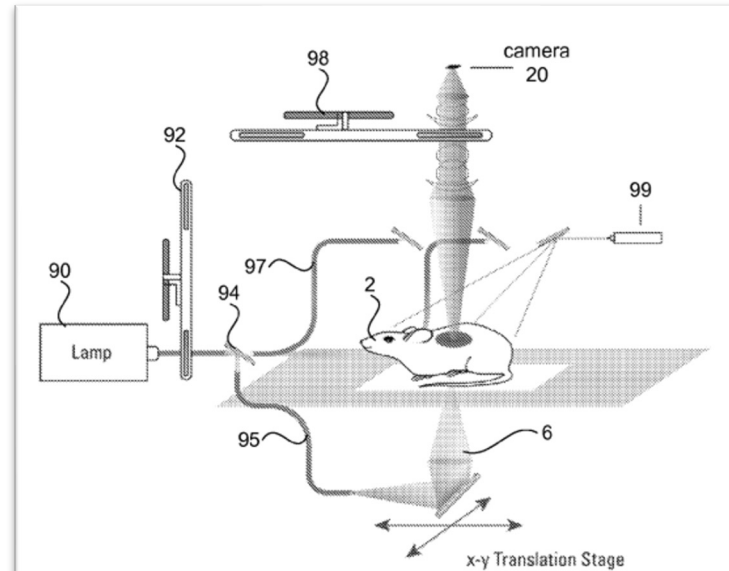
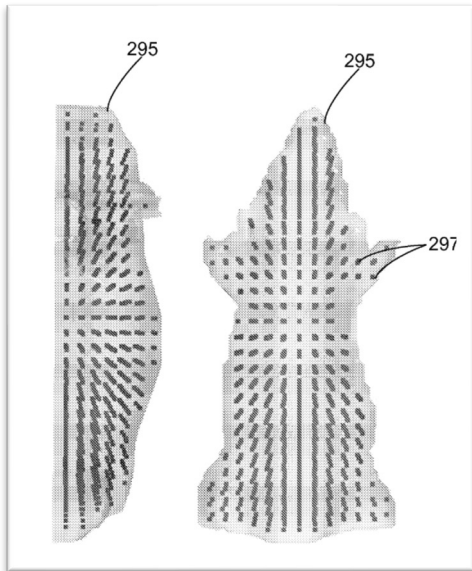


Figure 6A

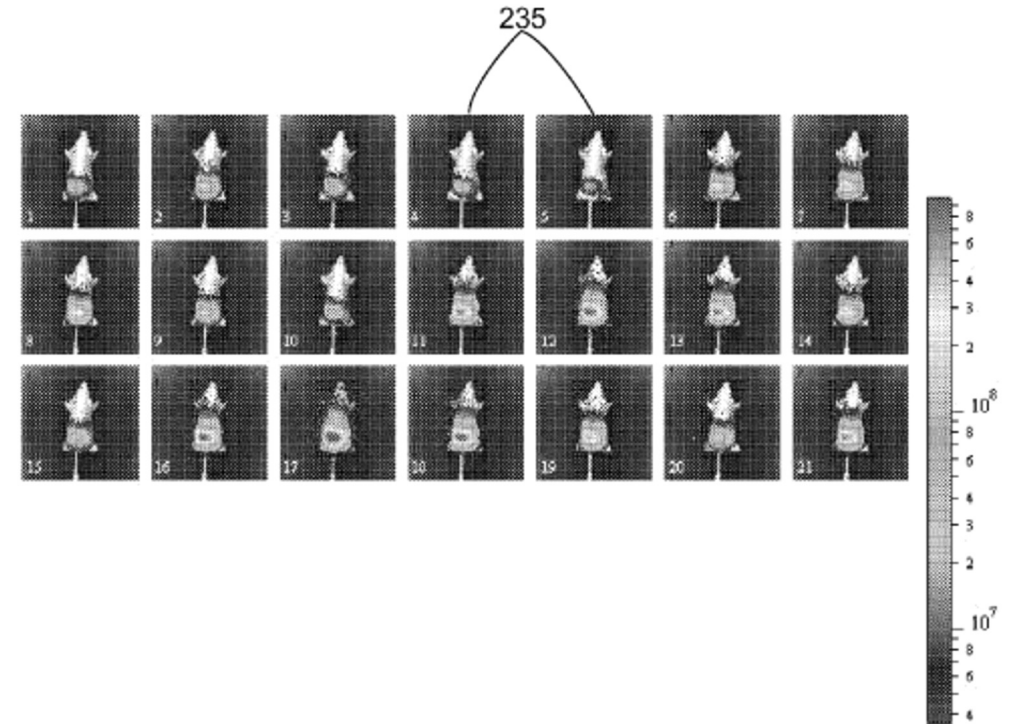
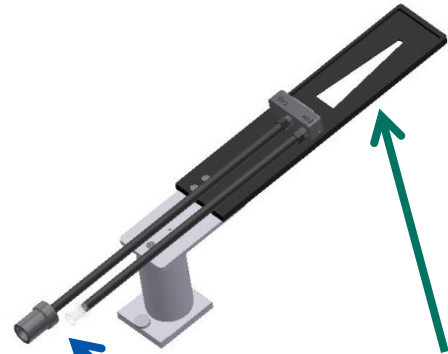


Figure 7

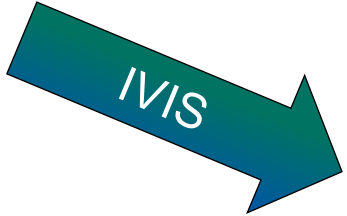
Mouse Imaging Shuttle for Co-Registration

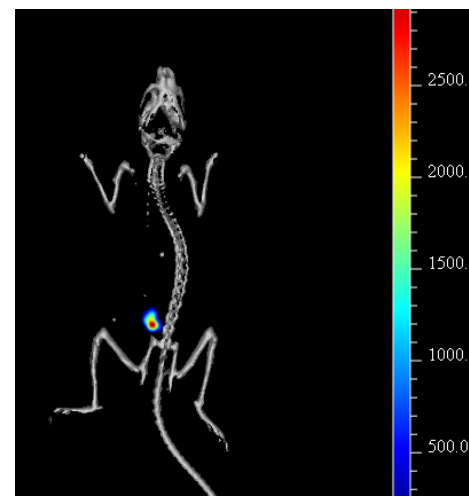
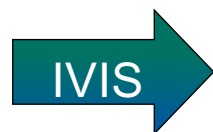
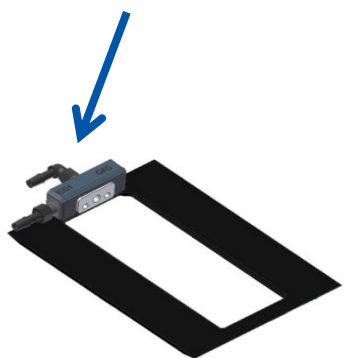
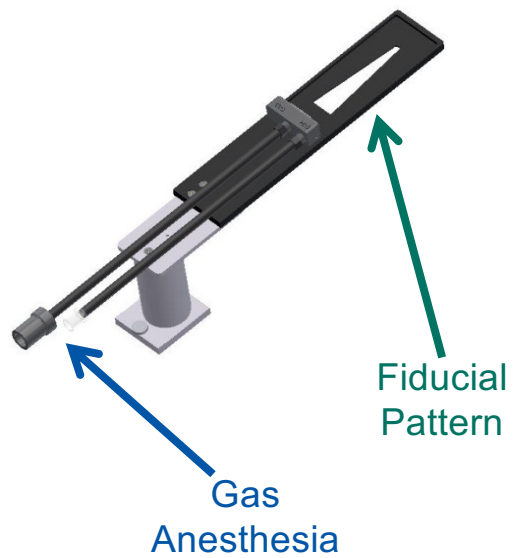
Shuttle Heights:
10, 12, 14 & 25 mm



Fiducial
Pattern

Gas
Anesthesia

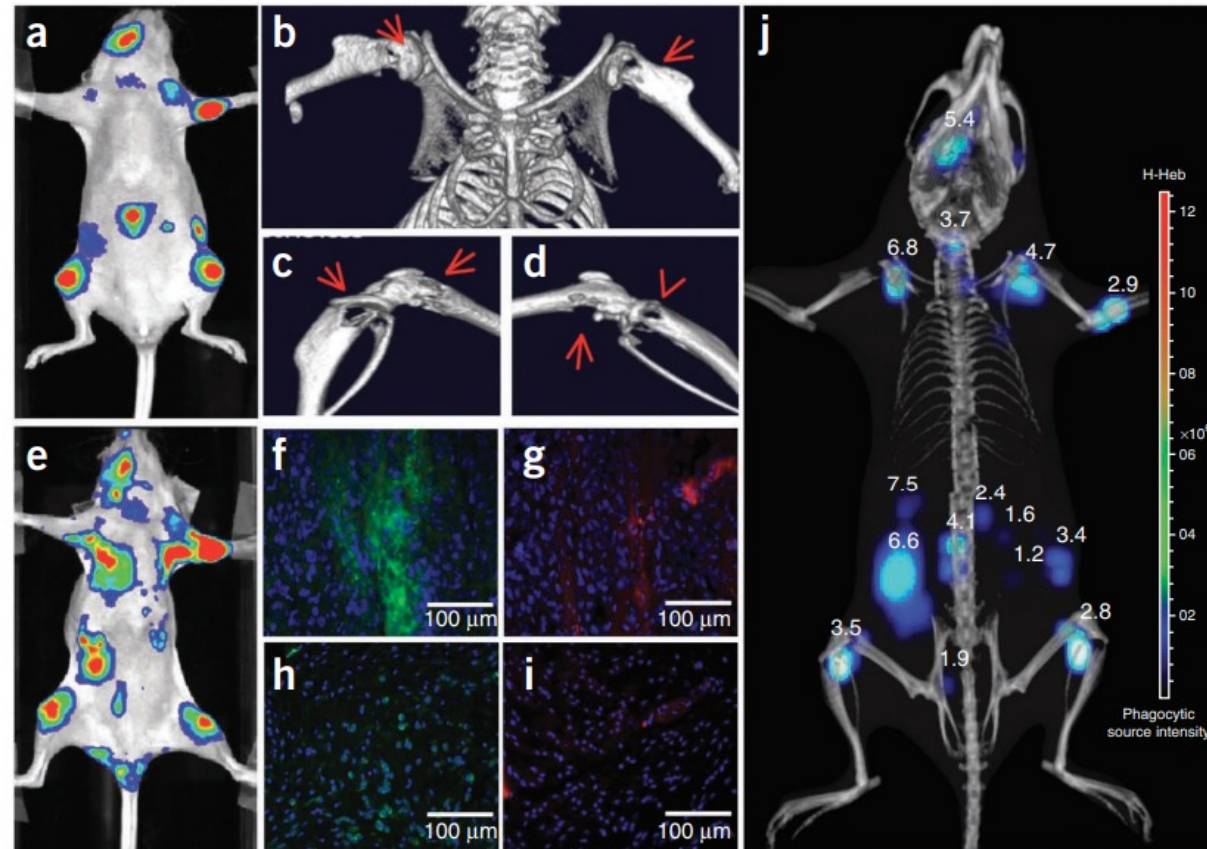




Enhanced detection of myeloperoxidase activity in deep tissues through luminescent excitation of near-infrared nanoparticles

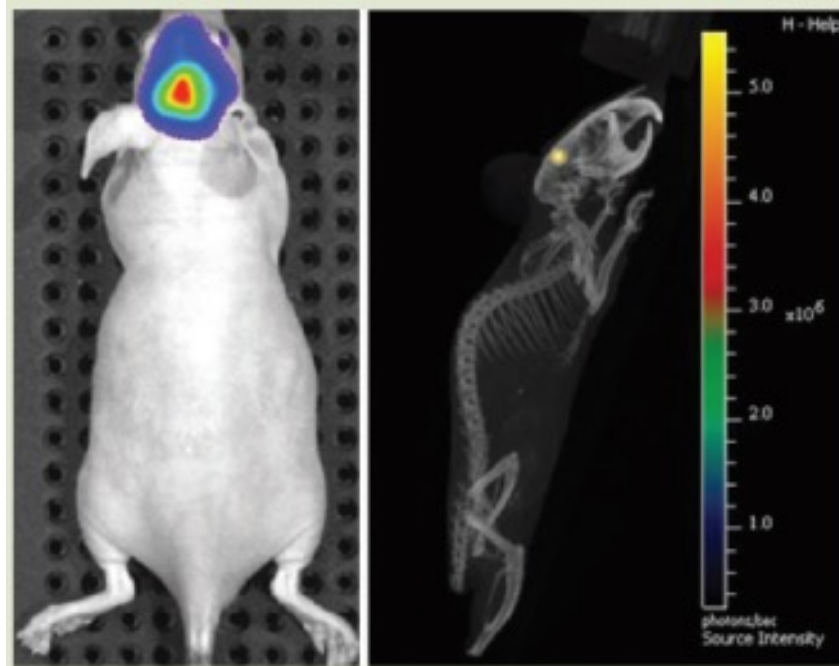
Ning Zhang^{1,3}, Kevin P Francis¹, Arun Prakash² & Daniel Ansaldi¹

Figure 5 Imaging MPO activity in MDA-MB-231-luc2 tumor metastases. **(a)** Bioluminescent imaging visualizing tumor metastases at 3 weeks in NU/NU mice intracardially injected with MDA-MB-321-luc2 tumor cells. **(b–d)** High-magnification microCT images obtained using a Quantum FX μ CT system of the shoulders and knee joints showing osteolytic lesions caused by tumor metastases. Arrows indicate regions of osteolytic lesions. **(e)** MPO activity in these lesions analyzed with Luminol-R. **(f–i)** Immunohistological analysis of the tumor metastases **(f,g)** or healthy tissues from a similar area in a control mouse **(h,i)** analyzed for neutrophils (green; **f,h**) and MPO (red; **g,i**) with a Nuance multispectral CCD camera. Nuclei are shown in blue. Original magnification, $\times 40$. **(j)** Co-registration of multimodality three-dimensional bioluminescent and microCT images obtained using an IVIS Spectrum instrument and a Quantum FX μ CT system, respectively. The DLIT-measured depth (mm) of the tumor metastases is shown above each of the metastatic nodules.

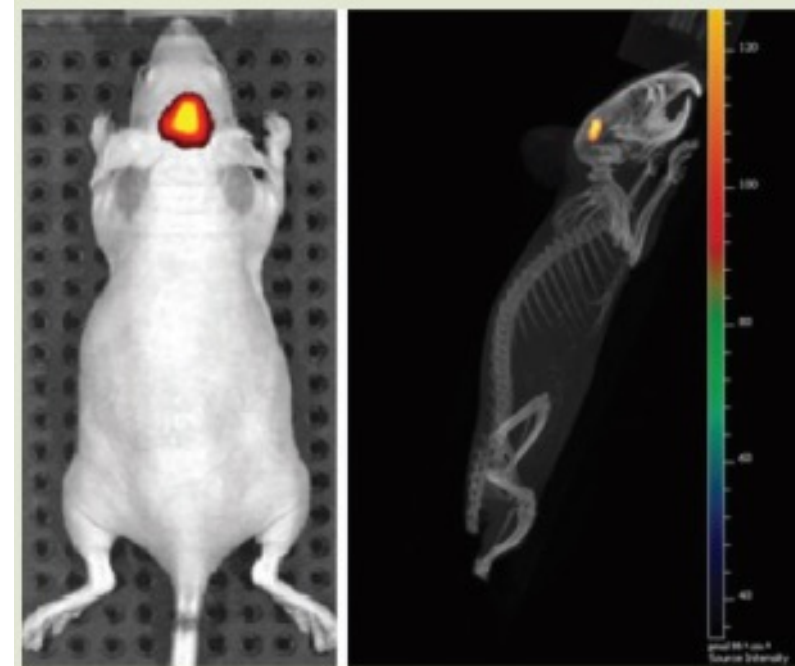


Multimodal Detection with Bioluminescent and Fluorescent Imaging Agents

- Bioware Brite U-87 MG-Red-Fluc orthotopic tumor cell (3×10^6) were implanted directly.
- DLIT and microCT were used in this study

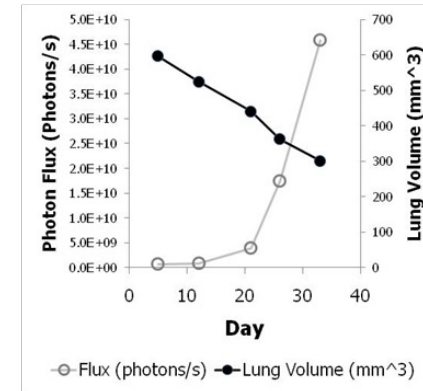
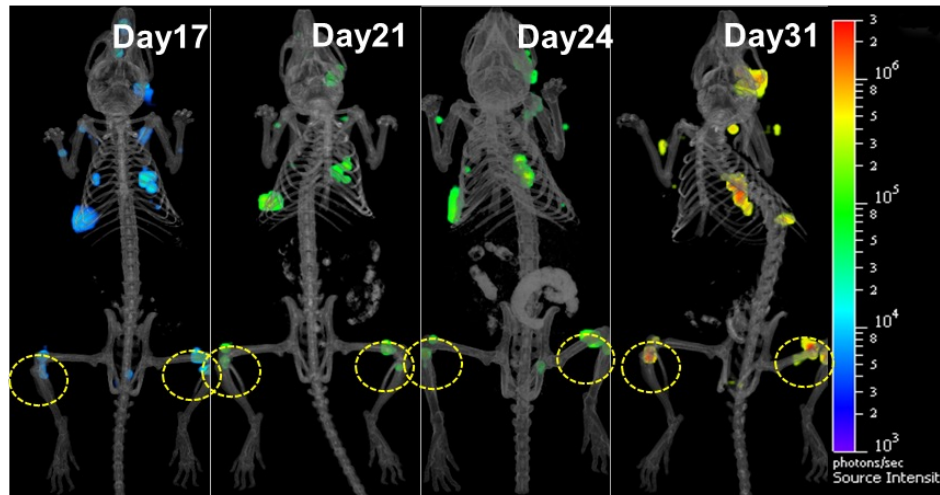
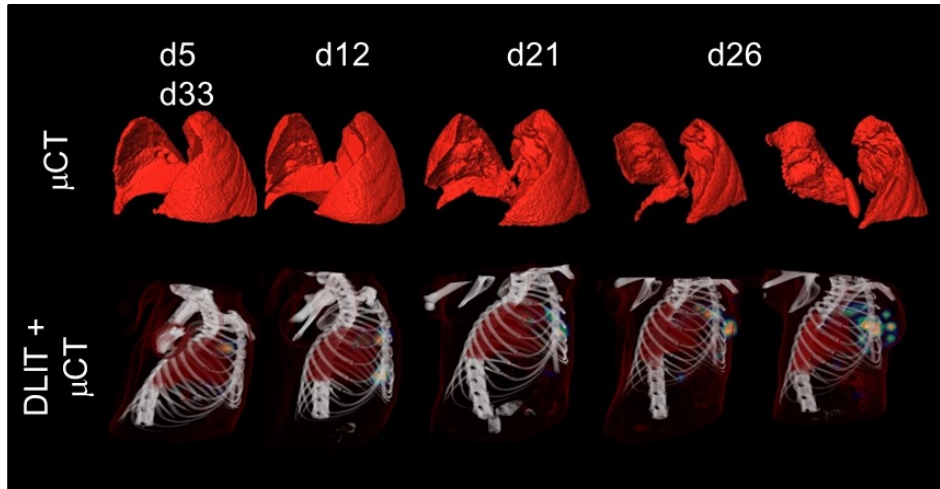


- ▶ Mouse was injected with a single dose of IntegriSense750 imaging agent to detect expression of integrin $\alpha v \beta 3$.
- ▶ FLIT and microCT were used in this study

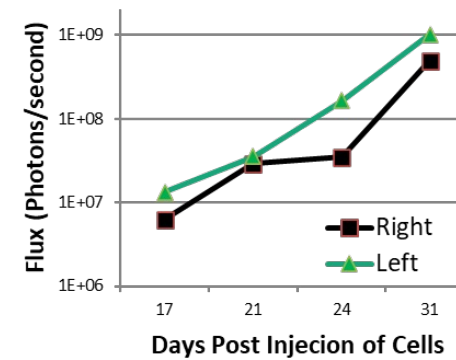


Composite functional and anatomical imaging obtained by using fluorescent and bioluminescent agents together gives a clearer context and understanding of the mechanisms of disease

Co-Registration Oncology Apps



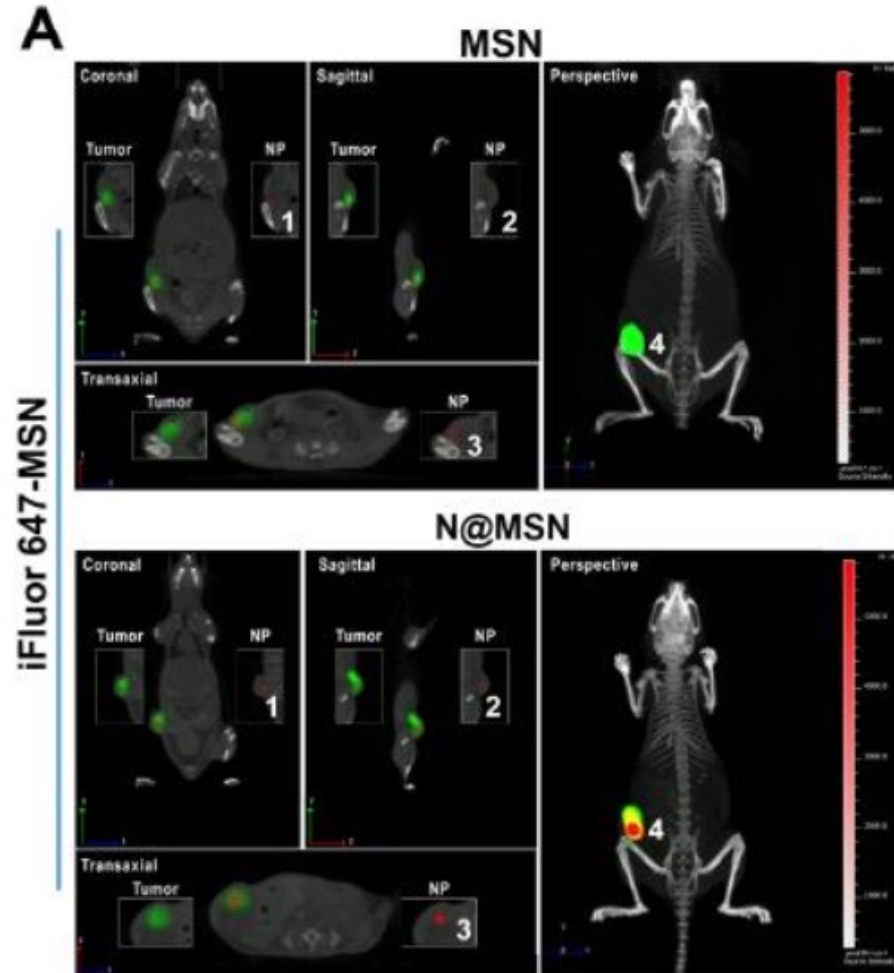
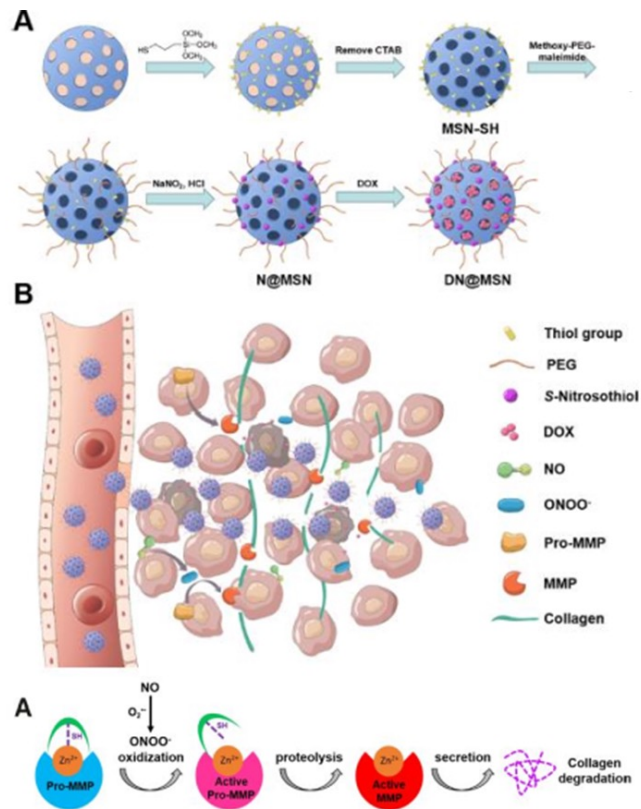
3D BLI Quantification of Metastases



REVIEW

Open Access

Near-infrared light-triggered nano-prodrug for cancer gas therapy



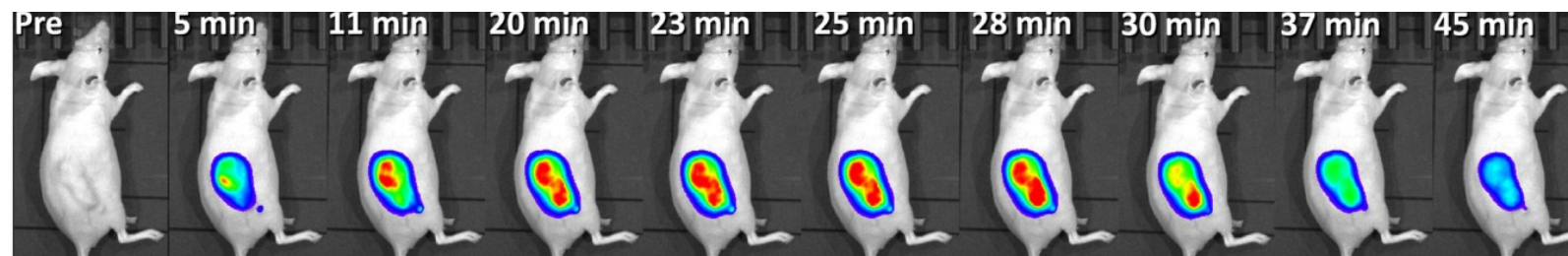
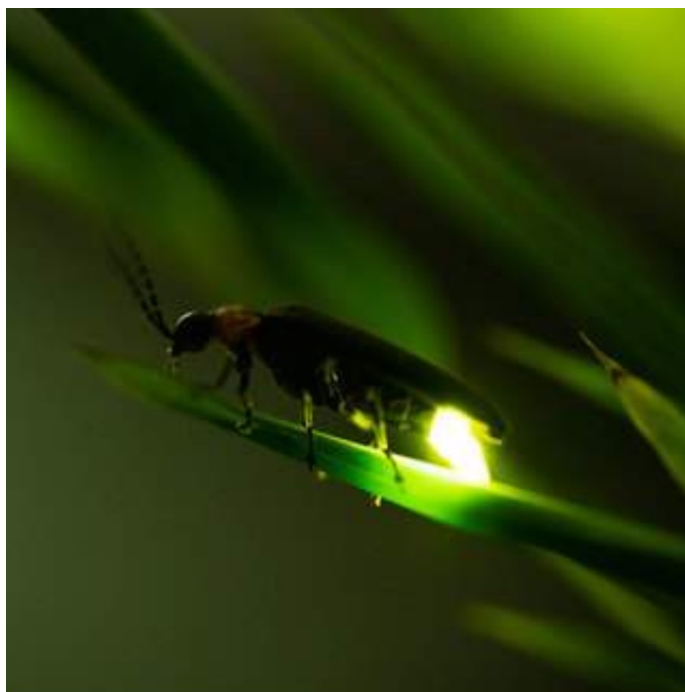
Top Ten Tips for Optical Imaging

1. Choose reporters that maximize signal-to-noise (S:N) ratio
2. Consider the appropriate control groups and imaging time points necessary
3. Use hairless mice or white-furred animals and depilate or shave
4. Switch to autofluorescence-free mouse diet
5. Closely map the kinetics of your biological bioluminescent model
6. Animal handling can significantly affect kinetics
7. Image in the animal orientation that yields the highest signal intensity
8. Cover intense signal to allow dimmer signals to dictate auto-exposure
9. Utilize guards to prevent reflection off neighboring animals
10. Use black well plates when doing in vitro experimentation



IVISbrite Bioluminescent Substrates and cell line

- RediJect™ solution – a pre-formulated, ready-to-use.
- Lyophilized D-Luciferin – offers the same sensitivity and high performance as the RediJect formulation.



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